

SESQUITERPENIC LACTONES OF *Inula aschersoniana* JANKA var. *aschersoniana* SPECIES*

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Received October 23, 1989

Accepted November 17, 1989

Aerial parts of species *Inula aschersoniana* JANKA var. *aschersoniana* have been shown to contain parthenolide (*I*), costunolide diepoxide (*III*) and the hitherto undescribed inusoniolide (*IV*) whose structure, including the absolute configuration, was determined using ¹H NMR and CD spectroscopy.

Within the framework of systematic investigation on sesquiterpenic lactones in species of the *Compositae* (*Asteraceae*) family we studied aerial parts of the species *Inula aschersoniana* JANKA var. *aschersoniana* (family *Compositae*, tribe *Inuleae*) which has been already shown to contain parthenolide¹ (*I*) and 8β-hydroxy-11α,13-dihydroeremanthin² (*II*). From this species we now isolated costunolide diepoxide (*III*), m.p. 163–164°C, of composition C₁₅H₂₀O₄. According to the ¹H NMR and IR spectra (see Experimental), the substance isolated was identical with the compound obtained earlier from species *Magnolia grandiflora* L. (family *Magnoliaceae*)³. This fact was proven by preparing costunolide diepoxide (*III*) from parthenolide (*I*) using the previously described procedure³. The synthetic diepoxide *III* was in all respects identical with the native compound.

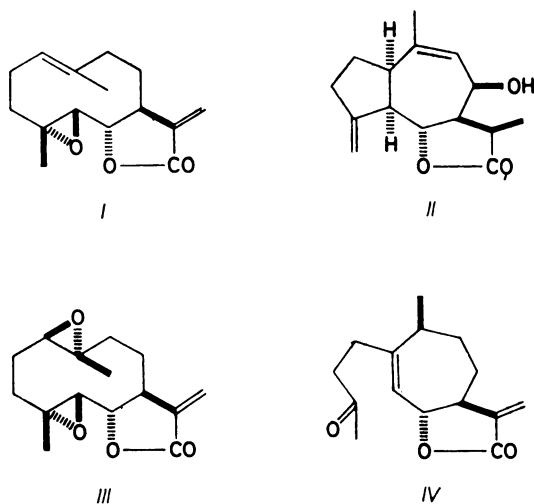
Another substance, which we have isolated from the described material, was a hitherto undescribed non-crystalline sesquiterpenic lactone *IV* of composition C₁₅H₂₀O₃, [α]_D –20.3°, which we named inusoniolide. Its infrared spectrum indicated the presence of a γ-lactone grouping whose carbonyl is conjugated with an exomethylene double bond (1 766, 1 406 and 1 160 cm⁻¹). The spectrum further revealed a keto group in an aliphatic chain or in a six-membered ring (1 713 cm⁻¹), and a double bond (1 666 cm⁻¹). The mass spectrum exhibited a molecular peak at

* Part CCCII in the series On Terpenes; Part CCCI: Collect. Czech. Chem. Commun. 55, 1059 (1990).

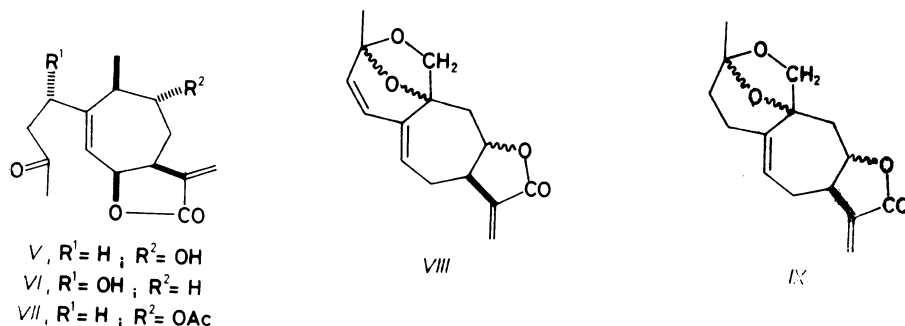
m/z 248 and the CD spectrum showed a Cotton effect at 245 nm ($\Delta\epsilon -0.5$) and at 200 nm ($\Delta\epsilon +9.5$). In the ^1H NMR spectrum (Table I) three proton signals proved the presence of a secondary methyl group (δ 1.09 d, $J = 7.5$ Hz) and a methyl ketone (δ 2.18 s). In the downfield part of the spectrum we found characteristic signals of an exomethylene group (δ 6.14 d, $J = 3.4$ Hz and δ 5.46 d, $J = 3.1$ Hz), a tri-substituted double bond proton (δ 5.68 m, $J = 2.8$ and 1.3 ($2\times$) Hz) and with it interacting CH—O lactone ring proton (δ 4.75 m, $J = 10.3$, 2.8 and 1.8 ($2\times$) Hz). The lactone ring thus must be closed in the position neighbouring with the double bond. The remaining ten protons in the molecule appeared as two non-analyzable multiplets in the regions δ 2.27–2.61 (6 H) and δ 1.64–1.86 (4 H). The ^{13}C NMR spectrum (Table I) confirmed the presence of the above-mentioned structural fragments ($\text{CH}_3\text{—CH}<$, $\text{CH}_3\text{—C(=O)—}$, $>\text{C=CH—}$ and an exomethylene- γ -lactone) and, moreover, proved four $\text{—CH}_2\text{—}$ groups. The degree of unsaturation, following from the elemental formula, together with the found structural fragments and the ^{13}C NMR spectrum, show unequivocally a bicyclic molecule. All these data enabled us to assign the 4,5-seco-guaianolide (xanthanolide) structure *IV* to inusoniolide.

TABLE I
 ^1H and ^{13}C NMR data of inusoniolide (*IV*) in CDCl_3

^1H NMR			^{13}C NMR	
Proton	δ	(J in Hz)	Carbon	δ
H-2	2.27–2.61 m		C-1	145.89 s
H-2'			C-2	33.01 t
H-3			C-3	41.56 t
H-3'			C-4	207.69 s
H-5	5.68 m ($J(5, 2) \approx J(5, 2') \approx J(5, 10) \approx 1.3$; $J(5, 6) = 2.8$)		C-5	123.63 d
			C-6	81.31 d
H-6	4.75 m ($J(6, 5) = 2.8$; $J(6, 7) = 10.3$; $J(6, 2) \approx J(6, 10) \approx 1.8$)		C-7	47.65 d
			C-8	23.82 t
H-7	2.27–2.61 m		C-9	30.73 t
H-8	1.64–1.86 m		C-10	37.40 d
H-8'			C-11	139.88 s
H-9			C-12	170.19 s
H-9'			C-13	118.65 t
H-10	2.27–2.61 m		C-14	15.64 q
H-13	6.16 d ($J(13, 7) = 3.4$)		C-15	29.90 q
H-13'	5.46 d ($J(13', 7) = 3.1$)			
H-14	1.09 d ($J(14, 10) = 7.5$)			
H-15	2.18 s			



The *trans*-annulation of the lactone ring follows from the allylic coupling constants $J(7, 13) = 3.4$ Hz and $J(7, 13') = 3.1$ Hz, the vicinal coupling constant $J(6, 7) = 10.3$ Hz and from the use of the so-called lactone rule⁴. In the hitherto described xanthanolides with γ -lactone ring closed at C(6) (for a review see refs^{5,6}) the lactone ring is invariably *cis*-annulated and the found values of the coupling constants ($J(6, 7) = 9-10$ Hz and $J(7, 13) \approx J(7, 13') = 2.5-3$ Hz) do not differ substantially from those found by us. However, there is a marked difference between the chemical shifts of the H-6 proton signals. Whereas in the spectra of the described *cis*-xanthan-6,12-olides the H-6 proton signal appears at about δ 5.50, in the spectrum of our compound *IV* this signal is located at δ 4.75. The upfield shift of the H-6 proton signal thus reflects the change in the annulation of the lactone ring from *cis* to *trans* and may be used as an important argument in favour of the *trans*-fusion in the lactone *IV*. Similar characteristic differences in the chemical shifts of the H-6 proton signal were observed by us for *cis*- and *trans*-2-oxo-1(10),3-dien-6,12-olides⁷ and we also discussed the reasons of this phenomenon. We have no direct evidence on the configuration of the methyl group at the C(10) carbon atom in compound *IV* and the tentatively suggested β -configuration is based on analogy with the structurally closest known compounds – ivalbatin⁸ (*V*) and parthemollin^{9,10} (*VI*) – in which the β -configuration of the C(10) methyl has been proven by X-ray diffraction. We also used comparison of the ¹³C NMR data of these compounds (or their acetates) for assignment of some signals in the ¹³C NMR spectrum of inusoniolide (*IV*). In this connection, it is worth notice that comparison of ¹H NMR spectra of acetyl-ivalbatin⁸ (*VII*) and fruticosin¹² (in which the position of the acetate group was not



determined) shows unequivocally that both these compounds are structurally identical.

The absolute configuration at C(7) in inusoniolide (IV) was derived from its CD spectrum. According to the Geissman's rule¹¹, the Cotton effect of the exomethylene- γ -lactone grouping at 245 nm ($\Delta\epsilon -0.5$), together with the *trans*-annulation of the γ -lactone ring closed at C(6) (as indicated by the ¹H NMR spectrum), corresponds to the *S*-configuration at the C(7) carbon atom.

The isolation of parthenolide (I) and costunolide diepoxide (III) agrees with the common occurrence of sesquiterpene germacrane lactones in the tribe *Inuleae* and particularly in the genus *Inula*^{5,6}. Xanthanolides (4,5-secoguaianolides) also occur in this taxa, although they are not so numerous as e.g. germacranolides or eudesmanolides^{5,6}. However, the described xanthanolides in both taxa mentioned have the γ -lactone ring closed on C(8), mostly with the β -configuration of the C(8)—O bond. Thus, inusoniolide is the first representative of xanthanolides from *Inula* genus and *Inuleae* tribe* and the first xanthanolide with *trans*-annulation of the γ -lactone ring to C(6).

EXPERIMENTAL

The melting points were determined on a Kofler block and they are not corrected. The IR spectra were measured in chloroform on a Perkin-Elmer PE 580 spectrophotometer. The ¹H and ¹³C NMR spectra (200 MHz or 50.3 MHz, respectively) were measured on a Varian XL-200 instrument in deuteriochloroform with tetramethylsilane (for ¹H) and deuteriochloroform (for ¹³C; $\delta(\text{CDCl}_3) = 77.0$) as internal reference. Chemical shifts and coupling constants of protons were obtained by first order analysis from the expanded spectra (2 Hz/cm) using exponential multiplication with gaussian apodization function for resolution enhancement. The multiplicity

* For some species of the *Inuleae* tribe, e.g. *Geigeria africana* (p. 372), *G. aspera* (p. 373) etc., griesenin (VIII) and its dihydro derivative IX are reported⁶ as xanthanolides with the γ -lactone ring closed at C(6). However, the structure of these xanthanolides clearly shows that the γ -lactone ring is closed at C(8).

of signals in ^{13}C NMR spectra was determined using the "attached proton test" pulse sequence¹³. The mass spectra were measured on an AEI MS 902 spectrometer. Optical rotation was determined on an objective polarimeter Perkin-Elmer 141 and CD spectra were measured on a Roussel-Jouan CD 185 dichrograph.

Isolation of sesquiterpene lactones. Dry aerial parts of the species *Inula aschersoniana* JANKA var. *schersoniana* STOY., STEF. et KITAN (*Compositae*) (voucher RL 286/76 is deposited in the herbarium of Zakład Roślin Leczniczych, Akademia Medyczna, Poznań, Poland) (3 kg) were worked up in the described manner¹⁴ and the obtained so-called lactone fraction (6 g) was chromatographed on a column of Kieselgel 60 (Merck, 500 g) in light petroleum-chloroform (1 : 1) and then with chloroform. The first fractions afforded non-crystalline inusoniolide (*IV*, 2.5 g); $[\alpha]_{\text{D}} -20.3^{\circ}$ (*c* 0.37, pyridine). IR spectrum (cm^{-1}): 1 766, 1 406, 1 160 (methylene- γ -lactone); 1 713, 1 252, 1 265 (aliphatic ketone); 1 666 (double bond). Mass spectrum, *m/z*: 248 (M), 230 (M-18), 190 (M-58). CD spectrum (nm, $\Delta\epsilon$): 245, -0.5 ; 200, $+9.5$ (last reading). For the ^1H and ^{13}C NMR data see Table I. For $\text{C}_{15}\text{H}_{20}\text{O}_3$ (248.3) calculated: 72.53% C, 8.12% H; found: 72.25% C, 7.94% H.

Further fractions were subjected to preparative thin-layer chromatography on Silufol in chloroform and afforded parthenolide (*I*), $\text{C}_{15}\text{H}_{20}\text{O}_3$, m.p. 109–110°C; $[\alpha]_{\text{D}} +65^{\circ}$ (*c* 0.1, methanol), which showed no depression of mixture melting point with an authentic specimen.

Still further fractions, on column chromatography on silica gel followed by preparative thin-layer chromatography on Silufol, afforded diepoxycostunolide (*III*, 150 mg), $\text{C}_{15}\text{H}_{20}\text{O}_4$, m.p. 163–164°C; $[\alpha]_{\text{D}} -30^{\circ}$ (*c* 0.1, methanol); no m.p. depression on admixture with costunolide diepoxide, prepared from parthenolide by a described procedure³. IR spectrum (cm^{-1}): 1 770 (C=O, γ -lactone); 1 663, 1 621 (C=C). Mass spectrum, *m/z*: 264 (M), 246 (M-18), 235 (M-CHO), 217 (M-CHO-18). ^1H NMR spectrum: 1.35 d, 3 H (H-14, *J*(14,9) = 9.6); 1.40 s, 3 H (H-15); 2.74 m, 1 H (H-7, *J*(7,6) = 9.2; *J*(7,8) = 7.2; *J*(7,8') = 1.7; *J*(7,13) = 3.6; *J*(7,13') = 3.3); 2.86 dd, 1 H (H-1, *J*(1,2) = 9.9; *J*(1,2') = 1.7); 2.91 d, 1 H (H-5, *J*(5,6) = 8.6); 3.95 t, 1 H (H-6, *J*(6,5) = 8.6; *J*(6,7) = 9.2); 5.63 d, 1 H (H-13', *J*(13',7) = 3.3); 6.35 d, 1 H (H-13, *J*(13,7) = 3.6).

Elemental analyses were performed in the Analytical Department of this Institute (Dr V. Pechanec, Head) by Mrs E. Listiková and Mr V. Štěřba. Optical rotations were determined by Mrs Z. Ledvinová. Mass spectra were measured and interpreted by Dr J. Kohoutová and Dr L. Dolejš. The IR and CD spectra were measured and interpreted by Dr S. Vašičková. We express our thanks to all of them as well as to Mrs M. Snopková for technical assistance. Some of us (E. B., W. M. D. and B. D.) thank the Polish Academy of Sciences, represented by its Institute of Organic Chemistry, for coordinating and financing a part of this study (problem 6.7: C.P.B.P. 01.13.2.20).

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Translated by M. Tichý.